



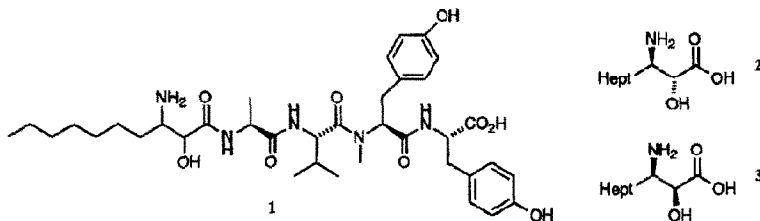
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Asymmetric Synthesis of (2*S*,3*R*)-3-Amino-2-Hydroxydecanoic Acid: The Unknown Amino Acid Component of Microginin

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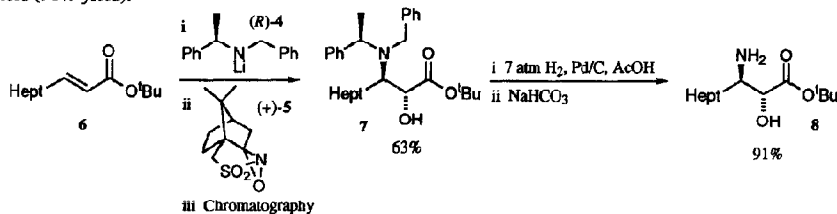
Abstract: 3-Amino-2-hydroxydecanoic acid (AHDA) is an unusual amino acid purported to occur in the recently isolated angiotensin-converting enzyme inhibitor microginin. In order to elucidate the stereochemistry of the naturally occurring material, and thus complete the structural assignment of microginin, both the (2*R*,3*R*)-*anti*-diastereoisomer and the (2*S*,3*R*)-*syn*-diastereoisomer of AHDA have been prepared. Comparison of the ¹H and ¹³C nmr spectroscopic data of the synthetic amino acids with that reported for the naturally occurring material indicates that the *relative* stereochemistry of the AHDA found in microginin is *syn*. The *absolute* stereochemistry of the natural amino acid is shown to be (2*S*,3*R*) by comparison of its reported CD spectrum with that recorded for the synthetic material prepared herein.

Microginin 1 is a pseudopentapeptidic angiotensin-converting enzyme (ACE) inhibitor which has recently been isolated from the freshwater blue-green alga *Microcystis aeruginosa*.¹ A number of techniques, including 2D nmr and chemical degradation, were employed to elucidate the partial structure of 1 with 3-amino-2-hydroxydecanoic acid (AHDA) being implicated as the *N*-terminal component. The structure of this novel β-amino-α-hydroxy amino acid was suggested by the ¹H and ¹³C nmr spectra, and the FAB mass spectrum, of a sample isolated by hydrolysis of microginin.¹ On the basis of a Cotton effect in the CD spectrum, it was further suggested that the α-stereogenic centre in this acid was of the *R* configuration, however this must be regarded as tentative, especially since the configuration of the β-stereogenic centre remained undefined.¹ In order to complete the structural and stereochemical assignment of microginin, this communication describes the asymmetric synthesis of authentic samples of the homochiral *anti* and *syn* diastereoisomers (2*R*,3*R*)-AHDA 2 and (2*S*,3*R*)-AHDA 3 for spectroscopic comparison with the naturally occurring acid.



We have previously shown that the conjugate addition of lithium (*R*)-(α -methylbenzyl)benzylamide (*R*)-4 to *tert*-butyl cinnamate, followed by *in situ* hydroxylation with (+)-(camphorsulfonyl)oxaziridine (+)-5, provides the corresponding *anti* β -amino- α -hydroxy amino acid derivative with excellent diastereoselectivity (92% d.e.).² We anticipated therefore that the corresponding reaction using the readily available³ enoate 6 would provide the basis for an efficient synthesis of 2. Indeed, the tandem addition-hydroxylation of 6 using (*R*)-4 and (+)-5 provided the desired adduct 7 with good diastereoselectivity (88% d.e.) and this material was

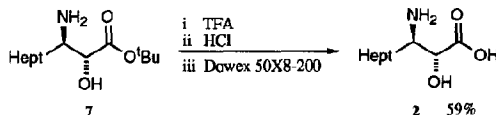
isolated as a single diastereoisomer in 63% yield after flash chromatography. Subsequent catalytic debenzoylation of **7** occurred without complication to afford the free β -amino- α -hydroxy ester **8** in excellent yield (91% yield).



The *anti* relative stereochemistry within **8** was confirmed by conversion to the oxazolidinone **9** using carbonyl diimidazole (CDI). Analysis of the ring proton coupling constant in **9** ($J_{4,5} = 8.6$ Hz) confirmed the *cis* stereochemistry^{4,5} which follows directly from the *anti* arrangement in **8**, and hence **7**. The *absolute* stereochemistry of **7** follows from addition of (*R*)-**4** to the *Re*-face of **6** in an analogous manner to that previously elucidated for a number of related enantiomers.^{2,5,6}



With the stereochemistry within **8** established, a convenient synthesis of the *anti* amino acid **2** was available. Thus, treatment of **8** with trifluoroacetic acid (TFA) to remove the *tert*-butyl group, conversion of the resultant TFA salt to the corresponding hydrochloride, and ion-exchange chromatography afforded the free amino acid **2** in 59% isolated yield: $[\alpha]_D^{25} +3.4$ (c 0.70, 1N HCl).



Analysis of the ¹H nmr and ¹³C nmr spectra of synthetic **2** was completely consistent with the designated structure but a comparison with the spectroscopic data reported^{1,7} for the naturally occurring acid (see tables) revealed sufficient inconsistency to suggest that **2** was not the component of microginin. Consequently, it appeared that the naturally occurring amino acid was of the *syn* relative stereochemistry and, in order to unequivocally confirm this assignment, a synthesis of (2*S*,3*R*)-AHDA **3** was undertaken.

We anticipated that **3** could be readily prepared from the *anti*- β -amino- α -hydroxy ester **8** using our previously reported oxazoline inversion protocol.² Thus, the *N*-benzoyl derivative **10** was prepared by treatment of **8** with one equivalent of benzoyl chloride and readily isolated in 83% yield. Intramolecular cyclisation of **10** under Mitsunobu conditions, with attendant stereochemical inversion at the α -centre, subsequently furnished the *trans*-oxazoline **11** ($J_{4,5} = 6.3$ Hz) in good yield (84%). Finally, exhaustive hydrolysis of **11** with 6N HCl, and ion exchange chromatography of the resultant hydrochloride salt, afforded

the desired free amino acid **3** as a single diastereoisomer in excellent yield (92%): $[\alpha]_D^{25} +5.4$ (*c* 0.59, 1*N* HCl). As anticipated, the ^1H nmr and ^{13}C nmr spectra of synthetic homochiral **3** correlated with the data recorded¹ for the natural acid (see tables) and confirmed that the unusual amino acid component found in microginin was indeed 3-amino-2-hydroxy-decanoic acid with *syn* relative stereochemistry.

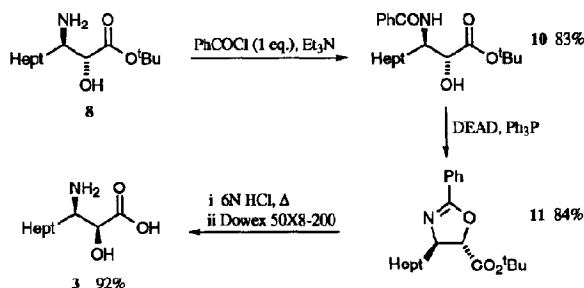


Table 1: ^1H nmr data for *anti* and *syn* 3-amino-2-hydroxydecanoic acid **2** and **3**

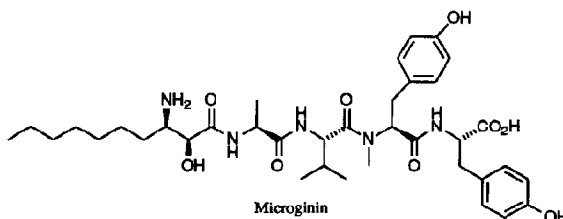
^1H nmr (d_6 -DMSO)	2	3	Natural Acid ¹
CH(OH)CHN	δ 3.35, d, <i>J</i> 8.1 Hz	δ 3.55, br	δ 3.54, m
CH(OH)CHN	δ 2.82, m	δ 3.10, br	δ 3.08, br
CH ₂ (CH ₂) ₅	δ 1.71–1.26, m	δ 1.60–1.20, m	δ 1.3–1.2, m
CH ₃ (CH ₂) ₅	δ 0.86, t, <i>J</i> 6.9 Hz	δ 0.86, t, <i>J</i> 6.8 Hz	δ 0.85, t

Table 2: ^{13}C nmr data for *anti* and *syn* 3-amino-2-hydroxydecanoic acid **2** and **3**

^{13}C nmr (d_6 -DMSO)	2	3	Natural Acid ¹
CO ₂	-	δ 173.4	-
CH(OH)CHN	δ 70.3	δ 69.5	δ 69.60
CH(OH)CHN	δ 53.9	δ 53.0	δ 52.30
CH ₃ (CH ₂) ₅	δ 31.3	δ 31.4	δ 31.17
	δ 30.2	δ 29.1	δ 28.90
	δ 29.0	δ 28.8	δ 28.90
	δ 28.6	δ 28.7	δ 28.48
	δ 24.8	δ 25.3	δ 25.20
	δ 22.1	δ 22.3	δ 22.07
CH ₃ (CH ₂) ₅	δ 14.0	δ 14.2	δ 13.95

It has been reported¹ that the CD spectrum of the AHDA isolated from microginin exhibited a negative Cotton effect at 215nm and, presumably on the assumption that the β -configuration was irrelevant, this was suggested to implicate the *R* stereochemistry at the α -stereogenic centre. However, the CD spectrum for synthetic **3** also indicated a negative Cotton effect in the same region⁸ (216nm) indicating that the previous stereochemical assumption was erroneous and that, in fact, **3**, with the (2*S*,3*R*) configuration, is the unknown amino acid component of microginin.⁹

In conclusion, the unknown amino acid component of microginin has been established as (2*S*,3*R*)-3-amino-2-hydroxydecanoic acid **3** by an independent asymmetric synthesis and spectroscopic comparison with data reported for a sample secured by hydrolysis of the natural product. These studies complete the structural and stereochemical assignment of microginin, for which the revised structure is depicted below.



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References

1. T. Okino, H. Matsuda, M. Murakami and K. Yamaguchi, *Tetrahedron Lett.*, 1993, **34**, 501.
2. M. E. Bunnage, S. G. Davies and C. J. Goodwin, *J. Chem. Soc., Perkin Trans. 1*, 1993, 1375.
3. J. M. Hawkins and T. A. Lewis, *J. Org. Chem.*, 1992, **57**, 2114.
4. S. Futagawa, T. Inui and T. Shiba, *Bull. Chem. Soc. Jpn.*, 1973, **46**, 3308.
5. M. E. Bunnage, S. G. Davies and C. J. Goodwin, *Synlett*, 1993, 731.
6. S. G. Davies and O. Ichihara, *Tetrahedron: Asymmetry*, 1991, **2**, 183.
7. We thank Dr. Murakami for advising us of the solvent (d^6 -DMSO) and instruments (600MHz ^1H nmr; 75MHz ^{13}C nmr) employed in the characterisation of the naturally occurring acid. The nmr spectra of the synthetic amino acids **2** and **3** reported herein were recorded using a Bruker AM500 spectrometer (500MHz ^1H nmr; 125MHz ^{13}C nmr).
8. The *anti* diastereoisomer **2** also exhibited a negative Cotton effect but at significantly lower wavelength (204 nm).
9. Dr. Murakami has recently informed us that he has independently reassessed the configuration of the 3-amino-2-hydroxydecanoic acid found in microginin also as (2*S*,3*R*).

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